

Variation in the nutritional composition of farmed Atlantic salmon (*Salmo salar* L.) fillets with emphasis on EPA and DHA contents

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Abbreviations

n-3 LC-PUFA (omega-3 long-chain polyunsaturated fatty acids); EPA (eicosapentaenoic acid); DHA (docosahexaenoic acid); 18:1n-9 (oleic acid); 18:2n-6 (linoleic acid) 18:3n-3 (α -linolenic acid); AOCS (American Oil Chemist's Society); EFSA (European Food Safety Authority); ISSFAL (International Society for the Study of Fatty Acids and Lipids); GOED (Global Organization for EPA and DHA Omega-3s); Se (selenium)

Declarations of interest: none

Abstract

The increase in the global popularity and production of farmed Atlantic salmon (*Salmo salar* L.) has led to compositional changes in their feeds that can potentially diminish their nutritive value. Thus, the aim of the study was to compare the lipid, protein, fatty acid (omega-3) and mineral contents of salmon fillet portions available in the UK and estimate their contribution towards consumer dietary intake levels. Twenty pre-packaged fresh salmon fillets, encompassing all ranges (value, standard, premium and organic) and farmed origins (Scotland and Norway) were purchased from 10 main UK-wide retailers and analysed for their nutritional compositions. Lipid contents were between 11.2-16.3% wet weight (ww), except the Retailer 10 value product which was significantly lower due to a high proportion of tail pieces. No difference in protein contents (17.5-20.2% ww) were observed between fillets. However, fatty acid profiles showed marked variations between samples with marker fatty acids 18:1n-9 (24.3-42.0%), 18:2n-6 (8.3-15.1%) and 18:3n-3 (2.6-8.1%) reflecting the differing levels of vegetable oil inclusion and eicosapentaenoic and docosahexaenoic acids (EPA+DHA, 5.6-16.6%) indicating the level of marine oils included within salmon feeds. Consequently, EPA+DHA contents varied from 0.88 to 2.36 g EPA+DHA.130 g⁻¹ flesh ww, equivalent to supplying 26 to 67% of the recommended 3.5 g EPA+DHA weekly intake suggested for optimal cardiac health in adults. Similarly, selenium contents differed significantly between samples delivering between 13.9-55.5% and 17.3-69.3% of the 75 and 60 µg.day⁻¹ UK intake for males and females, respectively. Additionally, EPA+DHA and selenium contents were both affected by farmed origin, reflecting differences in production strategies of the two salmon producing nations. Overall, the study highlights the contrasting nutritional profiles of farmed salmon fillets available to consumers based on retailer requirements (healthy versus sustainable product) and how this can affect the recommended dietary intakes from a human nutrition perspective.

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51 **Keywords:** farmed Atlantic salmon; aquaculture, EPA+DHA, selenium; retailer, consumers

1. Introduction

Since the introduction of intensive aquaculture in the 1960's, Atlantic salmon (*Salmo salar* L.) has grown to become a global commodity. Worldwide production of this high-value species now exceeds 2 million metric tonnes with Norway, Chile, Scotland and Canada the main producers among others (EY, 2018). Accordingly, the increased availability and thus, affordability, of farmed salmon has made it a popular choice among seafood consumers. In the EU, salmon is the most consumed farmed species as well as being the third most consumed fish species overall (EUMOFA, 2018). Global health authorities widely recommend consuming fish on a regular basis as part of a healthy diet due to their rich source of nutrients including protein, minerals and other micronutrients (EFSA, 2005; SACN/COT, 2004; WHO, 2003). Moreover, as an oily fish species, farmed salmon supplies a high level of omega-3 (n-3) long-chain polyunsaturated fatty acids (LC-PUFA), particularly eicosapentaenoic (EPA; 20:5n-3) and docosahexaenoic (DHA; 22:6n-3) acids, compared to many other fish species (Sprague et al., 2016, 2017a). These beneficial n-3 LC-PUFA, and to some extent docosapentaenoic acid (DPA; 22:5n-3), are widely acknowledged as being important for human health and development including neural function and in reducing chronic illnesses such as cardiovascular and inflammatory diseases among others (Calder, 2014; Kaur et al., 2011). However, the continual increase in the global population together with rising demand for seafood, including salmon, has resulted in changes to feed compositions that can negatively affect the final nutritional quality of farmed fish products.

Traditionally, farmed salmon feeds relied upon the inclusion of the finite marine raw materials, fish oil and fishmeal. However, as the aquaculture industry has grown the natural source of these ingredients, i.e. wild capture fisheries, has stagnated resulting in increased substitution by alternatives of terrestrial plant-based origin (Ytrestøyl et al., 2015). Although salmon growth remains largely unaffected due to the nutritional requirements of fish still being

met, some of the nutrients delivered by farmed salmon that are important to human health have declined, especially EPA and DHA contents as well as some minerals and other micronutrients (Betancor et al., 2016; Sissener et al., 2013; Sprague et al., 2016), in addition to an associated decrease in some harmful elements such as heavy metals and organic pollutants (Nøstbakken et al., 2015). The UK salmon industry has, nevertheless, adapted to these challenges by creating a distinct market position for itself by supplying a differentiated product largely driven by retailers (Shepherd et al., 2017). Indeed, the UK grocery sector consists of several large retailers/supermarkets that cater for a diverse market ranging from budget/low-cost through to premium/high-end. In addition, some retailers also offer a variety of the same product in an attempt to market themselves towards specific consumer preferences or attitudes such as cost, nutrition, ethics and sustainability. One such product is organically-reared salmon where feed ingredients, and their inclusion levels, as well as rearing standards, may differ to conventionally-reared salmon (Lerfall et al., 2016). However, similar standards can also be applied to conventionally-reared salmon based on specific retailer requirements that could result in a product differing in terms of nutritional content from similar marketed products. As the largest consumers of fresh salmon in Europe, selling four times as much in terms of value and volume compared to salmon's nearest rival, Atlantic cod (*Gadhus morhua*) (EUMOFA, 2018; Seafish, 2018), UK retailers also import additional salmon products to meet consumer demand (Henriques et al., 2014; Seafish, 2018;). Consequently, from a consumer viewpoint the purchasing of fresh salmon products can be a potentially confusing experience due to the many different products and prices ranges available that may, or may not, indicate differences in nutritional quality.

While there has been extensive experimental research over the years on the replacement of marine ingredients with alternatives and their potential implications on flesh quality (Bell et al., 2001, 2004; Betancor et al., 2018; Kousoulaki et al., 2015; Sprague et al., 2015; Torstensen

et al., 2004; Turchini et al., 2009, 2011), these studies are not necessarily reflective of current commercial practices. Moreover, the availability, quality and cost of raw ingredients are constantly altering and will ultimately dictate the type and levels of ingredients used in feeds by individual producers as well as determining the final flesh quality. As such, there is a lack of information on the ‘actual’ levels of nutrients in farmed salmon, which is important with regards to human health since nutritionists and public health bodies rely upon food databases being accurate when establishing and reviewing dietary guidelines to ensure adequate nutrient intakes (De Roos et al., 2017; Elmadfa and Meyer, 2010; Merchant and Dehghan, 2006). Therefore, the present study sought to analyse and compare the nutritional composition, in terms of lipid, protein, fatty acid (EPA+DHA) and mineral contents, of all available farmed salmon fillets (i.e. value, standard, premium and organic ranges) sold within the main UK retailers and assess their contribution in supplying essential nutrients to the human consumer.

2. Materials and Methods

2.1. Sample collection and preparation

Fresh pre-packaged ‘own-brand labelled’ farmed Atlantic salmon fillet portions were purchased from the refrigerated section of 10 main UK retailers, with exception of one product where the retailer specialised in frozen products only. In addition, a brand-labelled salmon product, available from more than one retailer, was also purchased for comparison. Samples consisted only of raw fillet portions. Value-added products such as breaded or sauce-added products were excluded due to the potential interference of additional ingredients on nutrient profiles. All product ranges (value, standard, premium and organic), as defined by the retail product label, available at the time of study were purchased for analysis (see Table 1). If a retailer stocked only one product range, which was not organic, then this was deemed as their standard product, irrespective of the status of the retailer themselves. Products that shared the

same packaging, barcode identifier and nutritional labelling, but differed in farmed origin were treated as separate entities. Three samples per product range were purchased from UK-wide retailers within the Stirling and Clackmannan areas of Central Scotland. To minimise the potential of sampling salmon products from the same batch of fish, i.e. salmon farmed and harvested from the same cage/site, collection of replicate samples occurred over an eight-week period at four week intervals. Individual product packs were analysed as a single pooled sample, regardless of the number of portions contained in the pack (i.e. if a pack contained two portions, these were pooled to provide a single sample for analysis). Fillets were skinned and boned, where required, and homogenised on the day of purchase using a Robot-Coupe Blixer® 4 V.V. blender mixer (Robot-Coupe, Vincennes Cedex, France) before storing at -20°C until analysed. All analyses were performed on a wet weight (ww) basis, and in duplicate minimum, with a relative standard deviation of <5% between technical replicates deemed acceptable.

2.2. Lipid content and fatty acid analysis

Total lipid was extracted from ~0.5 g of homogenised salmon flesh in 20 volumes of ice cold chloroform/methanol (2:1, v/v) according to Folch et al. (1957). Non-lipid impurities were isolated by washing with 0.88% (w/v) KCl and the lipid weight determined gravimetrically following evaporation of solvent under oxygen-free nitrogen and overnight desiccation *in vacuo*.

Fatty acid methyl esters (FAME) were prepared from total lipid extracts by acid-catalysed transesterification at 50°C for 16 h using 2 mL of a 1% (v/v) solution of sulphuric acid (95%, Aristar®, VWR Chemicals, Poole, UK) in methanol and 1 mL of toluene (Christie, 1993). FAME were extracted and purified according to Tocher and Harvie (1988), based on the American Oil Chemist's Society (AOCS) methods for marine oils (Ce li-07 and Ce 1b-89; AOCS, 1992, 2007), and separated and quantified by gas-liquid chromatography (GC) using a

Fisons GC-8160 (Thermo Scientific, Milan, Italy) equipped with a 30 m × 0.32 mm i.d. × 0.25 µm ZB-wax column (Phenomenex, Cheshire, UK), 'on column' injection and flame ionisation detection. Hydrogen was used as carrier gas with the initial oven thermal gradient from 50°C to 150°C at 40°C.min⁻¹ to a final temperature of 230°C at 2°C.min⁻¹. Individual FAME were identified by comparison to known standards (Restek 20-FAME Marine Oil Standard, Thames Restek UK Ltd., Buckinghamshire, UK) and published data (Tocher and Harvie, 1988). Data were collected and processed using Chromcard for Windows (Version 1.19; Thermoquest Italia S.p.A., Milan, Italy). Fatty acid content per g of tissue was calculated using heptadecanoic acid (17:0) as internal standard (IS), which was included with the sample at a known concentration in order for fatty acid (FA) concentration to be calculated as $\text{Conc. FA (mg.g lipid}^{-1}) = \frac{\text{Peak Area FA} \times (\text{Conc. IS/Peak Area IS})}{\text{lipid value}}$ with the lipid value used to express the result on a ww sample basis (mg.g sample⁻¹).

2.3. Crude protein analysis

Crude protein was determined by weighing out ~0.25 g of sample and adding 5 mL sulphuric acid (analytical reagent grade, Fisher Scientific, Loughborough, UK) together with 2 copper catalyst tablets (Fisher Scientific, Loughborough, UK), before digesting at 400°C for 1 h (Foss Digester 2040; Foss Analytical AB, Högnäs, Sweden). Total nitrogen levels were measured by Kjeldahl (Foss Kjeltect™ 2300, Foss Analytical AB, Högnäs, Sweden) and the crude protein level calculated as $\text{N} \times 6.25$.

2.4. Minerals and trace elements

Total minerals and trace elements (calcium, cobalt, copper, iron, manganese, magnesium, phosphorus, selenium, sodium, vanadium and zinc) were determined using inductively coupled plasma mass spectrometry (ICP-MS) with collision cell technology (Thermo X, Series 2;

Thermo Scientific, Hemel Hempstead, UK) as previously outlined by Betancor et al. (2015). Briefly, 20-30 mg of sample was added to Teflon tubes along with 5 mL of 69% nitric acid (Aristar® analytical grade; VWR Chemicals, Poole, UK) and digested in a microwave digester (MARS Xpress; CEM Microwave Technology Ltd., Buckingham, UK) in three stages consisting of 21-190°C for 10 min at 800 W, then 190°C for 20 min at 800 W followed by a final 30 min cooling period. The digested solution was transferred into 10 mL volumetric flasks and made up to volume with deionised water before 0.4 mL of this solution was transferred to 10 mL centrifuge tubes and made up to volume with deionised water before analysing by ICP-MS. For total selenium determination, 10 µL internal standard (Gallium and Scandium, 10 ppm; BDH Chemicals Ltd., Poole, UK) and 0.2 mL methanol, to ensure sensitivity, was added to 0.4 mL of initial digest solution before making up to 10 mL with deionised water prior to analysis by ICP-MS. The ICP-MS operated in kinetic energy discrimination (KED) mode using 100 % helium as collision gas to correct for any interference. Argon was used as plasma gas. A certified reference material (Fish Muscle ERM-BB42; Institute for Reference Materials and Measurements (IRMM), Geel, Belgium) was included with sample batches to monitor the integrity of the sample procedure.

2.5. Quality assurance

The method of performance of the analytical procedures described above were further assessed through the satisfactory annual performance of interlaboratory proficiency test including: the European Federation for the Science and Technology of Lipids, organised by the German Society for Fat Science (DGF, Frankfurt, Germany), for fatty acid content parameters; Masterlab Analytical Services BV (Boxmeer, Netherlands) for analytical methods routinely used in the feed, oil, fish producing and technology sectors; and AOCS (Illinois, USA) for the fatty acid content of marine oils attaining Approved Chemist status.

2.6. Statistical analyses

Statistical analyses were performed using Minitab® v17.1.0 statistical software package (Minitab Inc., Pennsylvania, USA). Data were analysed for normality with Kolmogorov-Smirnov test and for homogeneity of variances by Bartlett's test together with the examination of residual plots and, where necessary, transformed by arcsine or natural logarithm. Data were compared by a one-way analysis of variance (ANOVA) with multiple comparisons made using Tukey's post hoc test. A significance of $P < 0.05$ was applied to all statistical tests performed. Significant differences between data in tables and figures are indicated by different superscript lettering. A principal component analysis (PCA) was used as the ordination method of farmed origin based on the parameters measured in the study in order to distinguish the farmed origin of the unknown sample product from Retailer 4. All data are presented as the mean and standard deviation (mean \pm sd).

3. Results and Discussion

3.1. Lipid and protein contents

The lipid and protein contents of the various farmed salmon products are presented in Supplementary Table 1. Lipid levels were generally in the range 11.2-16.3% wet weight (ww), consistent with those reported elsewhere for commercially farmed salmon (Henriques et al., 2014; Jensen et al., 2012; Nichols et al., 2014; Sprague et al., 2016). Only the value-based product from Retailer 10 contained a significantly lower lipid level ($6.9 \pm 2.4\%$) than most other salmon fillets. This was related to the high proportion of tail pieces found in this particular product range. Lipid levels are known to vary throughout the flesh fillet, both anteriorly-posteriorly and dorsally-ventrally, with higher contents found around the dorsal fin region and the lowest levels at the tail end (Bell et al., 1998; Katikou et al., 2001). Furthermore, as dietary

lipid is a major source of energy, high-energy (lipid) diets are fed to salmon to ensure optimal growth by sparing the more expensive dietary protein for conversion to muscle tissue. However, dietary lipid content also influences lipid deposition (Sargent et al., 2002) and, will therefore contribute to variation in flesh lipid contents. In contrast, protein content of flesh showed less variation between samples with levels ranging from 17.5-20.2% ww.

3.2. Fatty acid composition

The primary concern related to the replacement of marine fish oil in salmon feeds with vegetable oils of terrestrial origin is that these oils are devoid of EPA and DHA, and so their use has a significant impact on the nutritional value of farmed products. Flesh lipid fatty acid profiles, presented as proportions of total fatty acids, of the farmed salmon products surveyed in the present study exhibited a high degree of variation. In particular, oleic (18:1n-9), linoleic (18:2n-6) and α -linolenic (18:3n-3) acids, which are typically characteristic of vegetable oil inclusion (Sargent et al., 2002; Sprague et al., 2016), ranged from a low of 24.3, 8.3 and 2.6%, respectively, in the Retailer 8 Scottish premium product to a high of 42.0, 15.1 and 8.1%, respectively, in the Retailer 2 Norwegian standard product (Figure 1a-c). In contrast, the fatty acids characteristic of marine fish oil inclusion, EPA and DHA, were notably lower in those samples containing higher levels of 18:1n-9, 18:2n-6 and 18:3n-3 and vice versa (Figure 1d). Lipid in commercial aquafeeds is generally comprised of a blend of fish and vegetable oils to meet the nutritional requirements of the fish being farmed, as well as for economic and ecological factors that have arisen from the increased demand for seafood from a growing population. Since the fatty acid composition of fish flesh is primarily determined by diet (Sargent et al., 2002), the present study highlights the many different dietary oils, both source and inclusion level, currently used in feed formulations by the industry to deliver a differentiated supply of salmon products. This may affect consumer choice when purchasing

salmon products as the nutrient profiling of foods, such as fat content and fatty acid profiles, can be influential in deciding whether specific nutritional claims (e.g. low in saturated fat, high in omega-3 etc.) can be included on product labels (Lobstein and Davies, 2009). Given the development of farmed animal feeds, particularly salmon, where the type and inclusion level of ingredients are constantly changing according to availability and price (Sissener et al., 2013; Sprague et al. 2016; Ytrestøyl, et al., 2015), the potential impact of the various feed formulations on the final product nutritional quality should be acknowledged and subsequently incorporated into food databases so that they remain up-to-date and minimise errors when assessing dietary human nutrient intake levels (Merchant and Dehghan, 2006).

While there were clear differences between product ranges (e.g. value, standard, premium and organic), the two premium (Retailers 1 and 8), two value (Retailers 8 and 10) and two organic (Brand and Retailer 10) products exhibited similar fatty acid profiles within their respective product ranges, suggesting comparable dietary oil sources and/or inclusions levels employed within the feeds for these ranges. However, a wider range of fatty acid compositions was revealed among the standard range of products. This variation is most likely a reflection of retailer status/values and/or the unique selling point of the product. The UK grocery sector is highly diverse, consisting of high-end premium retailers through to discount chains. In addition, many retailers market themselves on unique selling points relating to specific issues including nutritional quality, ethical and responsible sourcing, and sustainability (Shepherd et al., 2017). Accordingly, many of the standard salmon products would be expected to align with the ethos of the individual retailers. Thus, products sold in retailers promoting health benefits (i.e. omega-3) are expected to have higher levels of EPA+DHA due to increased dietary fish oil inclusion whereas retailers who market sustainable products would most likely have lower levels of these fatty acids due to increased plant oil inclusion (Henriques et al., 2014; Sprague et al., 2016). In the present study, retailers were not ranked on their perceived status owing to

the potential ambiguity arising from personal subjectivity. Notwithstanding, the wide variation of fatty acid profiles among retailers observed in the present study was also noted previously (Henriques et al., 2014).

Three of the ten retailers surveyed in the present study, together with the Brand, sold salmon products that encompassed more than one range: Retailer 1, standard and premium; Retailer 8, value, standard and premium; Retailer 10, value standard and organic; Brand, standard and organic. Both the premium products from Retailers 1 and 8 contained a greater EPA+DHA content as a percentage of total lipid, than their standard products (~14 vs. 10%, respectively), although not significant. Similarly, the Brand organic product contained a higher EPA+DHA level than its standard fillet range (10 vs 7%, respectively), whereas levels were similar for the organic and standard products from Retailer 10, around 9.5% of total lipid. Lerfall et al. (2016) found that organically farmed Norwegian salmon contained a higher proportion of EPA+DHA in the flesh than their conventionally-reared counterparts. Conversely, Di Marco et al. (2017) found a higher level of n-3 LC-PUFA in conventionally farmed seabass (*Dicentrarchus labrax*) and gilthead sea bream (*Sparus aurata*) as compared to organic. Organically-reared fish can regularly command higher prices than conventionally farmed salmon (Ankamah-Yeboah et al., 2016; Olesen et al., 2010). However, this isn't necessarily a reflection of the higher level of high-priced fish oil included in diets, but rather a quality assurance measure ascribed from certified organisations such as the EU's organic aquaculture standards (Regulation 710/2009) (EU, 2009), relating to a specific specification and/or conditions that differentiate it from comparable products. Similarly, particular specifications are applied to other quality products including *Label Rouge* salmon, where a maximum of 16% lipid is permitted in the flesh and fish must be fed on diets composed exclusively of marine products, vegetable ingredients, vitamin, minerals and carotenoids together with product traceability, maximum stocking densities and other codes of good practice (Label Rouge, 2013). In the UK, it is the retailers

which primarily determine the nutritional quality of the farmed salmon, particularly with respect to EPA+DHA contents and thus dietary fish oil inclusion levels (Shepherd et al., 2017). As feed generally represents half of the total production costs, the types and levels of dietary ingredients will typically determine the final cost in supplying the product.

Contrary to what might be expected, the value product from Retailer 10 contained a higher proportion of EPA+DHA ($12.0 \pm 4.3\%$) than both the standard and organic products ($9.5 \pm 3.0\%$ and $9.6 \pm 0.8\%$, respectively), albeit not statistically significant. This could suggest that the value product fish were fed a diet higher in EPA+DHA levels than the organic and standard fish. However, as mentioned above, the value product from retailer 10 contained a high proportion of tail pieces that had a lower lipid content. As such, the tail portions would be expected to contain a lower level of the main storage lipid, triacylglycerol, and a higher level of structural phospholipids than fattier portions. Although fatty acid compositions of both triacylglycerols and phospholipids are affected by dietary oil composition (Ruiz-Lopez et al., 2015), phospholipids generally contain a higher level of EPA and, especially, DHA as these play a crucial role in the structure of membranes (Sargent et al., 2002). Consequently, tail portions will generally have higher relative proportions of EPA and DHA than portions from fattier parts of the fillet. Therefore, while fatty acid profiles can provide an insight into the different feed formulations (dietary oil sources and levels) used, in addition to influencing whether specific health claims can be attached to food products (Lobstein and Davies, 2009), it is important to take into consideration both the lipid content and fatty acid composition when determining the absolute content of fatty acids (i.e. g.100 g flesh⁻¹) available to the human consumer.

3.3. EPA+DHA content and farmed origin

Overall, the average content of EPA+DHA of farmed Atlantic salmon products available on UK retailers' shelves was $1.1 \pm 0.4 \text{ g} \cdot 100 \text{ g}^{-1}$ flesh ww. However, when separated, based on farmed origin, Scottish salmon were found to contain a significantly ($P < 0.05$) higher amount of EPA+DHA ($1.3 \pm 0.4 \text{ g} \cdot 100 \text{ g}^{-1}$ ww) than their Norwegian counterparts ($0.8 \pm 0.1 \text{ g} \cdot 100 \text{ g}^{-1}$ ww) (Figure 2). This compares to 1.36 and 1.15 g EPA+DHA per 100 g⁻¹ ww salmon flesh reported in 2015 for Scottish (Sprague et al., 2016) and Norwegian farmed salmon (NIFES, 2016), respectively. This disparity can be explained largely by differences in production strategies. For example, the Norwegian salmon industry is the global powerhouse of farmed salmon production supplying around 1.2 million tonnes in 2016 compared to just 163,000 tonnes in Scotland (EY, 2018; Scottish Government, 2018). Consequently, the Norwegian industry is more reliant upon the use of sustainable feeds as the amount of fish oil used in aquaculture, approximately 800,000 metric tonnes per annum (Shepherd and Bachis, 2014), is spread thinner throughout the growing sector. Indeed, levels of fish oil in Norwegian salmon feeds fell from 24 to 11% between 1990 and 2013 (Ytrestøyl et al., 2015). Although Scottish salmon farmers face the same challenges as their Norwegian colleagues, as evidenced by the decline in EPA+DHA levels in Scottish salmon farmed between 2006 and 2015 (Sprague et al., 2016), the smaller volume of salmon produced in Scotland has enabled the establishment of a niche market for the product. Therefore, inclusion of slightly higher levels of marine ingredients in feeds has resulted in a "quality" product aimed at a premium market (Shepherd et al., 2017). This is clearly illustrated by the data in Figure 1 in which Norwegian farmed salmon are generally clustered together and display higher levels of 18:1n-9, 18:2n-6 and 18:3n-3 as well as lower EPA+DHA levels, indicating a higher inclusion of vegetable oil use, than their Scottish equivalents. Even so, farmed Scottish salmon also presented a wide range of fatty acid profiles that gave varying EPA+DHA levels, providing further evidence that the Scottish sector delivers highly differentiated salmon products based on individual retailer

requirements. The importance of obtaining information with respect to the origin of food, and the substances found in food products, has previously been emphasised by many authors as this can greatly affect the usefulness of data from food composition tables when assessing nutrient intake levels (Elmadfa and Meyer, 2010; Merchant and Dehghan, 2006). However, while Norwegian feed formulations are made public (Ytrestøyl et al., 2015), Scottish data remain private either due to the vast amount of bespoke diets produced or more probably due to retailer confidentiality (Shepherd et al., 2017).

3.4. EPA+DHA content and the human consumer

As an oily fish, farmed salmon contributes greatly to the dietary intake of the beneficial EPA+DHA fatty acids by human consumers (Sprague et al., 2016, 2017a). Global health authorities generally recommend consuming at least one portion of oily fish per week as part of a healthy diet (EFSA 2005; SACN/COT, 2004; WHO 2003). Figure 3 shows the amounts of EPA+DHA in the various salmon fillets analysed in the present study based on a portion size of 130 g, as advised by the European Food Safety Authority (EFSA, 2005). Like fatty acid composition, EPA+DHA contents presented a high degree of variation ranging from 2.36 ± 0.85 to 0.88 ± 0.23 g.130 g flesh⁻¹ ww in the Retailer 1 premium product and the Retailer 10 value product, respectively. It is important that such variations are acknowledged, particularly by those compiling and using data from food composition tables, as they can affect the ability to provide an accurate basis for assessing dietary nutrient intake and influence whether policy makers need to revise dietary guidelines such as fish consumption (De Roos et al., 2017; Elmadfa and Meyer, 2010).

Significant differences were observed between the Retailer 1 premium product and all Norwegian farmed salmon products together with the standard and value products from Retailer 4 and 10, respectively. Similarly, the Retailer 6 standard product also contained a

376 significantly higher content of EPA+DHA than all Norwegian farmed products, except the
377 Brand product, as well as the standard and value products from Retailers 4 and 10, respectively.
378 Lipid content and fatty acid profiles (reflecting oil sources) of salmon flesh ultimately
379 determine the absolute amounts of EPA and DHA delivered to the human consumer. For
380 example, feeds formulated with an EPA+DHA content of either 5 or 7.5% of lipid would be
381 expected to deliver 0.96 and 1.32 g EPA+DHA.100 g flesh⁻¹ ww, respectively, based on a fillet
382 lipid level of 17.5% (Ewos, 2013). Comparing the EPA+DHA contents in the Retailer 9
383 Norwegian standard product and the Retailer 10 value product (0.8 ± 0.1 and 0.7 ± 0.2 g
384 EPA+DHA.100 g⁻¹, respectively) suggests that fish were fed similar diets. However, the lipid
385 content of the former was twice that of the latter product ($15.1 \pm 1.2\%$ compared to $6.9 \pm 2.4\%$,
386 respectively). Had they been fed identical formulated diets, the Retailer 9 Norwegian standard
387 product would have given a higher EPA+DHA content. Examination of the fatty acid profiles
388 reveal that EPA+DHA levels, as a proportion of the lipid, were 6.1 and 12.0% (Norwegian
389 standard Retailer 9 and value Retailer 10, respectively), indicating that the value product from
390 Retailer 10 was fed a diet containing a higher level of fish oil, and therefore more EPA+DHA
391 than the Norwegian product from Retailer 9. Thus, the low EPA+DHA content in the value
392 product from Retailer 10 can be explained by the low lipid content due to the high presence of
393 tail pieces. While a similar caveat can be applied to all the samples surveyed in the study, in
394 that it can often be difficult for a purchaser to distinguish where precisely on the fillet a portion
395 is derived with the exception of tail pieces, no other apparent unconformities were noted. As
396 previously highlighted, other than the tail portions from Retailer 10's value product, lipid
397 contents were typically within range for farmed salmon (Henriques et al., 2104; Jensen et al.,
398 2012; Nichols et al., 2014; Sprague et al., 2016). As flesh fatty acid profiles reflect that of the
399 dietary oil (Sargent et al., 2002), diet composition can be considered the main determinant of
400 the EPA+DHA content variation reported in the present study.

401 Considering salmon products of different ranges available from the same retailer, premium
402 products from Retailer 1 and 8 contained a higher, albeit not significant, amount of EPA+DHA
403 (2.36 ± 0.85 and 1.88 ± 0.28 g EPA+DHA.130 g flesh⁻¹ ww, respectively) than their respective
404 non-premium products (1.39 ± 0.11 g.130 g flesh⁻¹ ww Retailer 1 standard range, and $1.65 \pm$
405 0.23 and 1.59 ± 0.40 g.130 g flesh⁻¹ ww for Retailer 8 value and standard ranges, respectively)
406 suggesting differences in feed formulations, and hence fish oil inclusion level, between
407 premium and other 'lower-value' product ranges. In contrast, Retailer 10's standard product
408 had higher EPA+DHA (1.46 ± 0.75 g.130 flesh⁻¹ ww) than both organic and value ranges (1.34
409 ± 0.21 and 0.88 ± 0.23 g.130 flesh⁻¹ ww, respectively) whereas, the Brand organic and standard
410 products contained more similar EPA+DHA contents of 1.28 ± 0.19 and 1.19 ± 0.18 g.130 g
411 flesh⁻¹ ww, respectively. Interestingly, when EPA+DHA contents were expressed in terms of
412 the amount delivered versus product cost, both the organic products (Retailer 10 and Branded)
413 yielded the lowest amount of EPA+DHA (g) per GBP (£) spent (0.39 ± 0.06 and 0.45 ± 0.07 g
414 EPA+DHA.£⁻¹, respectively) (Figure 4). While organically-reared fish can be considered a
415 premium product, the price does not necessarily reflect the inclusion level of fish oil used in
416 the feeds, but takes into account provenance and rearing conditions among other criteria.
417 Consumers are willing to pay up to 15% more for ethically-labelled products such as organic
418 or Freedom Food, a strict animal welfare assurance scheme, than salmon that have been
419 conventionally-reared (Olesen et al., 2010). Certainly, in retailers where more than one product
420 range was available, an increase in the cost per kg was seen as the product range increased, e.g.
421 £11.00, £13.75 and £20.03 per kg for Retailer 8 value, standard and premium ranges,
422 respectively (Table 1). This could indicate that the higher price for premium products reflects
423 enhanced or stricter farming condition. For instance, the value product from Retailer 8
424 generally gave a higher, although not significant, amount of EPA+DHA per £ (1.15 ± 0.16
425 g.EPA+DHA.£⁻¹) than the standard and premium ranges (0.89 ± 0.22 and 0.69 ± 0.10

g.EPA+DHA.£⁻¹, respectively). In contrast, Retailer 1 premium product gave more value (EPA+DHA) for money (1.25 ± 0.45 g.EPA+DHA.£⁻¹) than the standard product (1.07 ± 0.09 g.EPA+DHA.£⁻¹), suggesting that this retailer included higher priced ingredients (i.e. fish oil) to supply a product with improved health benefits (i.e. EPA+DHA). This is supported by the fact that the premium product from this retailer contained more EPA+DHA per serving than the standard range (see above and Figure 3). For Retailer 10 however, all products gave a similar result (0.39-0.45 g.EPA+DHA.£⁻¹) together with being the lowest overall providers of EPA+DHA per £ spent. In order to distinguish themselves from their counterparts, retailers will try to establish a unique selling point by creating a diverse range of salmon products based on certain attributes such as health-benefits, sustainability and/or price (Shepherd et al., 2017). Thus, it is plausible that the ethical principles that Retailer 10 uphold, e.g. lower stocking densities and other welfare/rearing conditions and the use of high-quality ingredients in feeds, either alone or in combination contributed to the higher price of the products.

The reduced use of marine ingredients in aquafeeds and consequent effects on EPA+DHA levels in farmed fish (Sprague et al., 2016; Ytrestøyl et al., 2015), has resulted in questions over whether current dietary guidelines regarding fish intake are sufficient to benefit human health (De Roos et al., 2017). Although at present there is no global consensus on a recommended level for EPA and DHA intake for humans, several advisory bodies have put forward their own recommendations for intakes to promote human health. On average, none of the products sampled in the present study would satisfy the commonly accepted 3.5 g EPA+DHA weekly intake (500 mg.day^{-1}) suggested by the International Society for the Study of Fatty Acids and Lipids (ISSFAL) to support optimal cardiac health in adults and endorsed by the Global Organization for EPA and DHA Omega-3's (GOED, 2019). This is not surprising as Sprague et al. (2016) reported that, on average two servings of farmed salmon in 2015 would be required to supply 3.5 g EPA+DHA owing to the changes in fish oil inclusion in salmon

feeds. However, five of the products analysed in the present study would, nevertheless, fulfil the lower recommendation of 1.75 g EPA+DHA per week (250 mg.day⁻¹) advocated by EFSA (2005). Obviously, two portions of these products would therefore satisfy the higher intake levels recommended by ISSFAL and GOED. Notwithstanding, the wide variation in EPA+DHA contents of the farmed salmon fillets sampled in the present study, providing 26 to 67% of the 3.5 g weekly EPA+DHA recommendation, should alert nutritionists to the fact that EPA+DHA levels in farmed salmon can vary so markedly, based on different production strategies, subsequently affecting the amount consumed in order to meet the recommended nutrient requirement.

Irrespective of the disparities in dietary recommendations, farmed salmon has been shown to deliver more EPA+DHA to the consumer than most other seafood products (Sprague et al., 2016; 2017a). Furthermore, both Henriques et al. (2014) and Sprague et al. (2016, 2017a) showed that farmed salmon contained higher EPA+DHA contents than wild Pacific salmon (*Oncorhynchus* sp.), also available in retailers, whereas the EPA+DHA content of wild Atlantic salmon has been reported to vary between 0.8 g (Jensen et al., 2012) to around 1.6 g EPA+DHA.100 g flesh⁻¹ ww (Lundebye et al., 2017). While no minimum EPA+DHA level has been officially adopted for salmon feeds, a level of around 1 % of the feed (equivalent to 3 % of dietary oil fraction) has been reported to maintain growth in salmon (Rosenlund et al., 2016; Bou et al., 2017), although fish performance has been shown to be compromised under challenging conditions (Bou et al., 2017). At these low inclusion levels, and taking into account endogenous production, just 0.63 g EPA+DHA.100 g flesh⁻¹ ww would be expected (Rosenlund et al., 2016). Novel sources of n-3 LC-PUFA have been trialled in farmed salmon in a bid to halt and reverse the further decline of these beneficial fatty acids, including microalgae (Kousoulaki et al., 2015; Sprague et al., 2015) and transgenic oilseed crops (Betancor et al., 2016b, 2018), with some already being used or nearing commercialisation

(Sprague et al., 2017b; Tocher et al., 2019). However, both availability and price associated with costs in producing these sources will inevitably determine the extent to which they are used. Nevertheless, the nutritional value of farmed salmon products is not served by EPA+DHA alone but also includes other important micronutrients such as minerals and trace elements, especially selenium (Lund, 2013; Tocher, 2015).

3.5. Minerals and trace elements

Fish consumption provides a rich source of minerals and trace elements such as calcium, phosphorous, zinc and selenium to the human consumer. The mineral contents of samples were relatively conserved between the various salmon products, although some significant differences in macro minerals (calcium and sodium) and all trace elements (copper, iron, manganese, selenium, vanadium and zinc) measured were observed (Table 2). Essential minerals are generally supplemented in feeds, particularly given the low cost of inorganic minerals used in premixes, whereas others such as calcium may also be absorbed from seawater (Lall and Dumas, 2015). However, while the nutritional requirements of the farmed fish can be met (NRC, 2011), the levels of some important nutrients from a human consumer viewpoint have fallen due to ingredient changes in farmed fish feeds (Sissener et al., 2013). Indeed, selenium (Se) content was found to vary markedly between the different salmon products, ranging from 0.08 ± 0.03 to 0.32 ± 0.01 mg Se.kg flesh⁻¹ ww for the Retailer 8 standard and Brand organic products, respectively. Selenium is a trace element essential for human health, being an important component of the antioxidant enzyme glutathione peroxidase, as well as playing a key role in immune function and thyroid metabolism (BNF, 2001). Current UK recommended daily intakes (RDI) are set at 75 and 60 µg.day⁻¹ for males and females, respectively (PHE, 2016). Thus, one 130 g portion of the Brand organic salmon would provide 41.6 µg Se, equivalent to 55.5 and 69.3% of the RDI for males and females, respectively.

Conversely, a 130 g portion of Retailer 4's standard product would give 10.4 µg equating to just 13.9 and 17.3% of the RDI for males and females, respectively. These differences can frequently be overlooked by nutritionists, who often assume farmed salmon to be of equal nutritional value. However, basic understanding of why such differences can occur are crucial to establishing and accurately estimating nutrient intakes.

Fishmeals contain higher levels of available Se than plant-based meals such as soybean or corn gluten (Betancor et al., 2016a; Gabrielsen and Opstvedt, 1980). The levels in plant products are determined by the concentrations and availability of Se in the soil where they are grown. Consequently, Se concentrations vary from country to country and are generally low in the UK and Europe compared to the Se-rich soils in North America (Fordyce, 2005). Therefore, the levels of Se in salmon flesh are dependent upon both the ingredient type and, for plant-based material, where it was grown. Being aware that identical foodstuffs from different origins can differ in their nutrient content is critical when establishing food databases as it can minimise the errors associated with the estimation of human dietary intakes (Elmadfa and Meyer, 2010; Merchant and Dehghan, 2006). There was, however, a significant difference in Se content based on farmed origin with Scottish salmon containing a higher content on average than their Norwegian counterparts (0.17 ± 0.06 versus 0.11 ± 0.03 mg Se.kg⁻¹ ww, respectively). These differences are most likely related to the lower use of fishmeal in Norwegian salmon feeds (Ytrestøyl et al., 2015). The replacement of marine ingredients has been linked to the decline in Se and other nutrients such as iodine and vitamin D in Norwegian salmon feeds between 2000 and 2010 (Sissener et al., 2013). Furthermore, Betancor et al. (2016a) demonstrated that, by increasing the substitution of marine ingredients by terrestrial plant-products in salmon feeds, the amount of Se in the flesh available to consumers was reduced.

Highest Se contents were found in the two organic products (0.32 ± 0.01 and 0.23 ± 0.07 mg Se.kg⁻¹ ww Brand and Retailer 10, respectively), indicating either higher inclusion of fishmeal or the use of high-quality plant ingredients grown in Se-rich soils such as North American wheat. However, the standard product from Retailer 10 also contained a similar Se content (0.26 ± 0.06 mg.kg⁻¹ ww), whereas all other products, including other Scottish farmed salmon, contained less than 0.16 mg Se.kg⁻¹ ww. Nevertheless, the range of Se concentrations among Scottish farmed salmon (0.13-0.32 mg.Se.kg⁻¹ ww) further reflects the wide range of feed formulations used by the Scottish salmon sector to supply a diverse selection of products (Shepherd et al., 2017).

3.6. Identifying unknown origin and labelling issues

The provenance and traceability of farmed animal products are of increasing importance to producers, retailers and consumers alike. In the present study, the precise origin of the salmon product from Retailer 4 was unknown, being labelled as either “farmed in Scotland or Norway”. The fatty acid composition of this product could suggest that fish were of Norwegian origin as the profile was similar to other Norwegian farmed salmon products (e.g. Retailers 2, 7, 9 and Brand), containing higher 18:1n-9, 18:2n-6 and 18:3n-3 and lower EPA and DHA levels (see Figure 1). However, fatty acid profiles from Retailer 1 and 5 Scottish standard products also exhibited similar profiles to Norwegian fed fish. When PCA was applied, based on farmed origin and all measured parameters (i.e. lipid and protein contents, and fatty acid and mineral compositions), the output for Retailer 4 appeared to align more with salmon of Norwegian farmed origin which were generally tightly clustered, compared to Scottish farmed salmon which were more widely scattered (Figure 5). This further reflects the diverse range of products produced for the UK retail market produced by the smaller Scottish industry (Shepherd et al., 2017, see Section 3.3). In the present study, individual fillets originating from

the same packet were pooled but Henriques et al. (2014) observed marked differences in fatty acid compositions of salmon portions sold within the same packaging, implying that some retail suppliers may utilise fish from various farmed sources to fulfil supply demands.

The differential in feed composition, both within and between salmon producers, can pose problems from a retailers' perspective, particularly with respect to nutritional labelling. This is best illustrated in Figure 6 that compares products that had identical nutritional contents stated on the product label but were of different farmed origins. No significant difference ($P>0.05$) was observed between Scottish and Norwegian salmon from Retailer 9 (1.04 ± 0.27 and 0.81 ± 0.07 g EPA+DHA.100 g flesh⁻¹ ww, Scotland and Norway, respectively), suggesting that these fish were fed similar formulated feeds. Contrastingly, a significant difference ($P<0.05$) was observed between the two identically labelled products from Retailer 2 (1.40 ± 0.09 and 0.75 ± 0.15 g EPA+DHA.100g flesh⁻¹ ww, Scotland and Norway, respectively), which mirrored the respective average EPA+DHA contents of the two salmon producing nations (see Figure 2). Obviously, this difference in EPA+DHA content between these identically labelled products also resulted in a significant difference in the cost of the EPA+DHA (g delivered per £) (Figure 4). Furthermore, Retailer 2 also sold an additional Norwegian salmon product, containing 3 fillet portions compared to the standard 2 fillet portion packs, which had a different nutritional label attached (see Table 1). The analysed EPA+DHA content of this product (0.80 ± 0.27 g EPA+DHA.100 g flesh⁻¹ ww) matched the labelled content of 0.8 g EPA+DHA.100 g flesh⁻¹ ww and was comparable to the other (2 fillet) Norwegian product from Retailer 2, indicating that the fish used for these products were likely fed similar diets. Labelling is frequently used to educate consumers and assist them in making healthier choices (Elmadfa and Meyer, 2010). Nevertheless, it is important to appreciate that several factors can affect the nutritional content of salmon flesh throughout the production cycle. In addition to feed composition (i.e. lipid source and level) and where portions are taken on the fillet (Bell et al., 1998; Katikou et al.,

2001), fish body size can affect lipid status (Shearer, 1994), although final harvest weights of UK sold salmon generally tend to be within 3-5 kg range. Therefore, changes to production schedules such as early harvest of fish (with lower weight) will also affect the nutritional value of the products to the consumer. Salmon producers routinely monitor several nutritional parameters and, if necessary, alter feed programmes to attain targeted levels. However, the present study represents a true reflection of the nutritional content of salmon portions, encompassing all areas of the fillet.

4. Conclusions

In summary, the results from the current study demonstrate that marked variations occur in the nutritional content of farmed salmon fillets available in the UK, particularly with respect to fatty acid profiles, EPA+DHA and selenium contents. Consequently, these disparities have a knock-on effect on the actual amounts delivered to consumers and will therefore affect an individual's ability to meet their recommended nutrient intakes. No clear indicators were seen between product ranges (value, standard, premium or organic) or retailers, being most likely due to the combination of the unique selling point of the product range or the retailer themselves (e.g. healthy versus sustainable product). However, EPA+DHA and selenium were affected by farmed origin reflecting differences in production strategies between the two salmon farming nations. Nevertheless, farmed salmon still delivers a high, but variable amount of beneficial nutrients (i.e. n-3 LC-PUFA, selenium) to consumers. Therefore, deviations in the nutritional contents of farmed animal products, as evidenced in the present study with farmed salmon, necessitate further monitoring in order to ensure that nutritional databases remain updated.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Conflicts of Interest

The authors declare no conflict of interest.

References

Ankamah-Yeboah, I., Nielsen, M., Nielsen, R. (2016). Price premium of organic salmon in Danish retail sale. *Ecological Economics*, 122, 54-60.

AOCS (1992). Official Method Ce 1b-89, Fatty acid composition of marine oils by GLC. In *Official Methods and Recommended Practices of the AOCS*, Illinois, USA.

AOCS (2007). Official Method Ce 1i-07, Determination of saturated, cis-monounsaturated and cis-polyunsaturated fatty acids in marine and other oils containing long-chain polyunsaturated fatty acids (PUFAs) by capillary GLC. In *Official Methods and Recommended Practices of the AOCS*, Illinois, USA.

Bell, J.G., Henderson, R.J., Tocher, D.R., Sargent, J.R. (2004). Replacement of dietary fish oil with increasing levels of linseed oil: modification of flesh fatty acid compositions in Atlantic salmon (*Salmo salar*) using a fish oil finishing diet. *Lipids*, 39, 223-232.

Bell, J.G., McEvoy, J., Tocher, D.R., McGhee, F., Campbell, P.J., Sargent, J.R. (2001). Replacement of fish oil with rapeseed oil in diets of Atlantic salmon (*Salmo salar*) affects tissue lipid compositions and hepatocyte fatty acid metabolism. *Journal of Nutrition*, 131, 1535-1543.

Bell, J.G., McEvoy, J., Webster, J.L., McGhee, F., Millar, R.M., Sargent, J.R. (1998). Flesh lipid and carotenoid composition of Scottish farmed Atlantic salmon (*Salmo salar*). *Journal of Agricultural and Food Chemistry*, 46, 119-127.

624 Betancor, M.B., Almailda-Pagán, P.F., Sprague, M., Hernández, A., Tocher, D.R. (2015). Roles
625 of selenoprotein antioxidant protection in zebrafish, *Danio rerio*, subjected to dietary
626 oxidative stress. *Fish Physiology and Biochemistry*, 41, 705-720.

627 Betancor, M.B., Dam, T.M., Walton, J., Morken, T., Campbell, P.J., Tocher, D.R. (2016a).
628 Modulation of selenium tissue distribution and selenoprotein expression in Atlantic salmon
629 (*Salmo salar* L.) fed diets with graded levels of plant ingredients. *British Journal of*
630 *Nutrition*, 115, 1325-1338.

631 Betancor, M.B., Li, K., Bucerzan, V.S., Sprague, M., Sayanova, O., Usher, S., Han, L.,
632 Norambuena, F., Torrisen, O., Napier, J.A., Tocher, D.R., Olsen, R.E. (2018). Oil from
633 transgenic *Camelina sativa* containing over 25% n-3 long-chain PUFA as the major lipid
634 source in feed for Atlantic salmon (*Salmo salar*). *British Journal of Nutrition*, 119, 1378-
635 1392.

636 Betancor, M.B., Sprague, M., Sayanova, O., Usher, S., Metochis, C., Campbell, P.J., Napier,
637 J.A., Tocher, D.R. (2016b). Nutritional evaluation of an EPA+DHA transgenic *Camelina*
638 *sativa* in feeds for post-smolt Atlantic salmon (*Salmo salar* L.). *PloS one*, 11(7), e0159934.

639 BNF. (2001). *Selenium and Health*. London, UK: The British Nutrition Foundation.

640 Bou, M., Berge, G.M., Baeverfjord, G., Sigholt, T., Østbye, T.-K., Ruyter, B. (2017). Low
641 levels of very-long-chain n-3 PUFA in Atlantic salmon (*Salmo salar*) diet reduce fish
642 robustness under challenging conditions in sea cages. *Journal of Nutritional Science*, 6, e32.

643 Calder, P.C. (2014). Very long chain omega-3 (n-3) fatty acids and human health. *European*
644 *Journal of Lipid Science and Technology*, 116, 1280-1300.

645 Christie, W.W. (1993). Preparation of derivatives of fatty acids for chromatographic analysis.
646 In W.W Christie (Ed.), *Advances in Lipid Methodology Two* (pp. 69-111), Dundee, UK: The
647 Oily Press.

648 De Roos, B., Sneddon, A.A., Sprague, M., Horgan, G.W., Brouwer, I.A. (2017). The potential
649 impact of compositional changes in farmed fish on its health-giving properties: is it time to
650 reconsider current dietary recommendations? *Public Health Nutrition*, 20, 2042-2049.

651 Di Marco, P., Petochi, T., Marino, G., Priori, A., Finoia, M.G., Tomassetti, P., Porrello, S.,
652 Giorgi, G., Lupi, P., Bonelli, A., Parisi, G., Poli, B.M. (2017). Insights into organic farming
653 of European sea bass *Dicentrarchus labrax* and gilthead sea bream *Sparus aurata* through
654 the assessment of environmental impact, growth performance, fish welfare and product
655 quality. *Aquaculture*, 471, 92-105.

656 EFSA. (2005). Opinion of the scientific panel on contaminants in the food chain on a request
657 from the European Parliament related to the safety assessment of wild and farmed fish.
658 *EFSA Journal*, 236, 1-118.

659 Elmadfa, I. Meyer, A.L. (2010). Importance of food composition data to nutrition and public
660 health. *European Journal of Clinical Nutrition*, 64, S4-S7.

661 EU. (2009). Commission Regulation (EC) No 710/2009 of 5 August 2009 amending
662 Regulation (EC) No 889/2008 laying down detailed rules for the implementation of Council
663 Regulation (EC) No 834/2007, as regards laying down detailed rules on organic aquaculture
664 animal and seaweed production. Official Journal of the European Union, L204/15-34.
665 Retrieved on March 8, 2019 from: [https://eur-lex.europa.eu/legal-](https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32009R0710&from=EN)
666 [content/EN/TXT/PDF/?uri=CELEX:32009R0710&from=EN](https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32009R0710&from=EN)

667 EUMOFA. 2018. The EU Fish Market, 2018 Edition. Retrieved on February 15, 2019 from:
668 https://www.eumofa.eu/documents/20178/132648/EN_The+EU+fish+market+2018.pdf

669 EWOS (2013). Fish and marine omega-3 in salmon feed. *Spotlight*, pp. 1-15.

670 EY. (2018). The Norwegian aquaculture analysis 2017. Retrieved on February 15, 2019 from:
671 https://www.ey.com/Publication/vwLUAssets/EY_-

[The Norwegian Aquaculture Analysis 2017/\\$FILE/EY-Norwegian-Aquaculture-Analysis-2017.pdf](#)

Folch, J., Lees, M., Sloane Stanley, G.H. (1957). A simple method for the isolation and purification of total lipids from animal tissues. *Journal of Biological Chemistry*, 226, 497-509.

Fordyce, F. (2005). Selenium deficiency and toxicity in the environment, In O. Selinus (Ed.), *Essentials of medical geology: Impacts of the natural environment on public health* (pp. 373-415), New York, USA: Elsevier.

Gabrielsen, B.O., Opstvedt, J. (1980). Availability of selenium in fish meal in comparison with soybean meal, corn gluten meal and selenomethionine relative to selenium in sodiumselenite for restoring glutathione peroxidase activity in selenium-depleted chicks. *The Journal of Nutrition*, 110, 1096-1100.

GOED. (2019). Intake Recommendations. Retrieved on March 20, 2019 from: <https://goedomega3.com/intake-recommendations>

Henriques, J., Dick, J.R., Tocher, D.R., Bell, J.G. (2014). Nutritional quality of salmon products available from major retailers in the UK: content and composition of n-3 long-chain PUFA. *British Journal of Nutrition*, 112, 964-975.

ISSFAL (2004). Report of the sub-committee on: Recommendations for intake of polyunsaturated fatty acids in healthy adults, International Society for the Study of Fatty Acids and Lipids (ISSFAL). Retrieved on November 19, 2018 from: <https://www.issfal.org/assets/issfal%2003%20pufaintakereccomdfinalreport.pdf>

Jensen, I.J., Mæhre, H.K., Tømmerås, S., Eilertsen, K.E., Olsen, R.L., Elvevoll, E.O. (2012). Farmed Atlantic salmon (*Salmo salar* L.) is a good source of long chain omega-3 fatty acids. *Nutrition Bulletin*, 37, 25-29.

696 Katikou, P., Hughes, S.I., Robb, D. (2001). Lipid distribution within Atlantic salmon (*Salmo*
697 *salar*) fillets. *Aquaculture*, 202, 89-99.

698 Kaur, G., Cameron-Smith, D., Garg, M., Sinclair, A.J. (2011). Docosapentaenoic acid (22:5n-
699 3): A review of its biological effects. *Progress in Lipid Research*, 50, 28-34.

700 Kousoulaki, K., Østbye, T.-K.K., Krasnov, A., Torgersen, J.S. Mørkøre, T., Sweetman, J.
701 (2015). Metabolism health and fillet nutritional quality in Atlantic salmon (*Salmo salar*) fed
702 diets containing n-3 rich microalgae. *Journal of Nutritional Science*, 4 (e24), pp 13.

703 Label Rouge (2013). Exceptional quality. Retrieved on March 20, 2019 from;
704 <http://saumonecossais.com/en/label-rouge-scottish-salmon/exceptional-quality>

705 Lall, S.P., Dumas, A. (2015). Nutritional requirements of cultured fish: Formulating
706 nutritionally adequate feeds. In D. Allen Davies (Ed.), *Feed and Feeding Practices in*
707 *Aquaculture* (pp. 53-109), Cambridge, UK: Woodhead Publishing (Elsevier).

708 Lerfall, J., Bendiksen, E.Å., Olsen, J.V., Morrice, D., Østerlie, M. (2016). A comparative study
709 of organic- versus conventional farmed Atlantic salmon. I. Pigment and lipid content and
710 composition, and carotenoid stability in ice-stored fillets. *Aquaculture*, 451, 170-177.

711 Lobstein, T. Davies, S. (2009). Defining and labelling ‘healthy’ and unhealthy’ food. *Public*
712 *Health Nutrition*, 12, 331-340.

713 Lund, E.K. (2013). Health benefits of seafood: Is it just the fatty acids? *Food Chemistry*, 140,
714 413-420.

715 Lundebye, A.-K., Lock, E.-J., Rasinger, J.D., Nøstbakken, O.J., Hannisdal, R., Karlsbakk, E.,
716 Wennevik, V., Madhun, A.S., Madsen, L., Graff, I.E., Ørnsrud, R. (2017). Lower levels of
717 persistent organic pollutants, metals and the marine omega 3-fatty acid DHA in farmed
718 compared to wild Atlantic salmon (*Salmo salar*). *Environmental Research*, 155, 49-59.

719 Merchant, A.T., Dehghan, M. (2006). Food composition database development for between
720 country comparisons. *Nutrition Journal*, 5:2.

721 Olesen, I., Alfnes, F., Røra, M.B., Kolstad, K. (2010). Eliciting consumers' willingness to pay
722 for organic and welfare-labelled salmon in a non-hypothetical choice experiment. *Livestock*
723 *Science*, 127, 218-226.

724 Nichols, P.D., Glencross, B., Petrie, J.R., Singh, S.P. (2014). Readily available sources of long-
725 chain omega-3 oils: Is farmed Australian seafood a better source of the good oil than wild-
726 caught seafood? *Nutrients*, 6, 1063-1079.

727 NIFES (2016). Omega-3 fatty acids in Norwegian farmed salmon. Retrieved 12 December
728 2018 from: [https://nifes.hi.no/wp-](https://nifes.hi.no/wp-content/uploads/2016/12/omega3fattyacidsinnorwegianfarmedsalmon.pdf)
729 [content/uploads/2016/12/omega3fattyacidsinnorwegianfarmedsalmon.pdf](https://nifes.hi.no/wp-content/uploads/2016/12/omega3fattyacidsinnorwegianfarmedsalmon.pdf)

730 Nøstbakken, O.J., Hove, H.T., Duinker, A., Lundebye, A.-K., Berntssen, M.H.G., Hannisdal,
731 R., Lunestad, B.T., Maage, A., Madsen, L., Torstensen, B., Julshamn, K. (2015).
732 Contaminant levels in Norwegian farmed Atlantic salmon (*Salmo salar*) in the 13-year
733 period from 1999 to 2011. *Environment International*, 74, 274-280.

734 NRC. (2011). National Research Council (NRC). *Nutrient Requirements of Fish and Shrimp*.
735 The National Academies Press, Washington D.C., USA.

736 PHE (Public Health England) (2016). Government Dietary Recommendations: government
737 recommendations for energy and nutrients for males and females aged 1-18 years and 19+
738 years. Retrieved January 30, 2019 from:
739 [https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_d](https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/618167/government_dietary_recommendations.pdf)
740 [ata/file/618167/government_dietary_recommendations.pdf](https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/618167/government_dietary_recommendations.pdf)

741 Rosenlund, G., Torstensen, B.E., Stubhaug, I., Usman, N., Sissener, N. (2016). Atlantic salmon
742 require long-chain n-3 fatty acids for optimal growth throughout the seawater period.
743 *Journal of Nutritional Science*, e19.

744 Ruiz-Lopez, N., Stubhaug, I., Ipharraguerre, I., Rimbach, G., Menoyo, D. (2015). Positional
 745 distribution of fatty acids in triacylglycerols and phospholipids from fillets of Atlantic
 746 salmon (*Salmo salar*) fed vegetable and fish oil blends. *Marine Drugs*, 13, 4255-4269.
 747 Sargent, J.R., Tocher, D.R., Bell, J.G. (2002). The Lipids In J.E. Halver, & R. W. Hardy (Eds.).
 748 *Fish Nutrition* (3rd ed) (pp. 181-257). San Diego, California, USA: Elsevier (Academic
 749 Press).
 750 SCAN/COT. (2004). Scientific Advisory Committee on Nutrition and Committee on Toxicity
 751 (SACN/COT). *Advice on fish consumption: benefits and risks*. 204 pp. The Stationary
 752 Office, Norwich, UK.
 753 Scottish Government. (2018). Marine Scotland Science: Scottish Fish Farm Production Survey
 754 2017. Retrieved March 25, 2019 from:
 755 [https://www.gov.scot/binaries/content/documents/govscot/publications/statistics/2018/10/s](https://www.gov.scot/binaries/content/documents/govscot/publications/statistics/2018/10/scottish-fish-farm-production-survey-2017/documents/marine-scotland-science-scottish-shellfish-farm-production-survey-2/marine-scotland-science-scottish-shellfish-farm-production-survey-2/govscot%3Adocument?forceDownload=true)
 756 [cottish-fish-farm-production-survey-2017/documents/marine-scotland-science-scottish-](https://www.gov.scot/binaries/content/documents/govscot/publications/statistics/2018/10/scottish-fish-farm-production-survey-2017/documents/marine-scotland-science-scottish-shellfish-farm-production-survey-2/marine-scotland-science-scottish-shellfish-farm-production-survey-2/govscot%3Adocument?forceDownload=true)
 757 [shellfish-farm-production-survey-2/marine-scotland-science-scottish-shellfish-farm-](https://www.gov.scot/binaries/content/documents/govscot/publications/statistics/2018/10/scottish-fish-farm-production-survey-2017/documents/marine-scotland-science-scottish-shellfish-farm-production-survey-2/marine-scotland-science-scottish-shellfish-farm-production-survey-2/govscot%3Adocument?forceDownload=true)
 758 [production-survey-2/govscot%3Adocument?forceDownload=true](https://www.gov.scot/binaries/content/documents/govscot/publications/statistics/2018/10/scottish-fish-farm-production-survey-2017/documents/marine-scotland-science-scottish-shellfish-farm-production-survey-2/marine-scotland-science-scottish-shellfish-farm-production-survey-2/govscot%3Adocument?forceDownload=true)
 759 Seafish (2018). Market insight factsheet: Chilled seafood in multiple retail. Retrieved February
 760 15, 2019 from:
 761 https://www.seafish.org/media/publications/Chilled_Seafood_in_Multiple_Retail_2018.pdf
 762 [f](https://www.seafish.org/media/publications/Chilled_Seafood_in_Multiple_Retail_2018.pdf)
 763 Shearer, K.D. (1994). Factors affecting the proximate composition of cultured fishes with
 764 emphasis on salmonids. *Aquaculture*, 119, 63-88.
 765 Shepherd, C.J., Bachis, E. (2014). Changing supply and demand for fish oil. *Aquaculture*
 766 *Economics & Management*, 18, 395-416.

767 Shepherd, C.J., Monroig, O., Tocher, D.R. (2017). Future availability of raw materials for
768 salmon feeds and supply chain implications: The case of Scottish farmed salmon.
769 *Aquaculture*, 467, 49-62.

770 Sissener, H., Julshamn, K., Espe, M., Lunestad, B.T., Hemre, G.I., Waagbo, R. Mage, A.
771 (2013). Surveillance of selected nutrients, additives and undesirables in commercial
772 Norwegian fish feeds in the years 2000-2010. *Aquaculture Nutrition*, 19, 555-572.

773 Sprague, M., Betancor, M., Dick, J, Tocher, D. (2017a). Nutritional evaluation of seafood with
774 respect to long-chain omega-3 fatty acids, available to UK consumers. *Proceedings of the*
775 *Nutrition Society*, 76(OCE2).

776 Sprague, M, Betancor, M.B., Tocher, D.R. (2017b). Microbial and genetically engineered oils
777 as replacements for fish oil in aquaculture feeds. *Biotechnology Letters*, 39, 1599-1609.

778 Sprague, M., Dick, J.R., Tocher, D.R. (2016). Impact of sustainable feeds on omega-3 long-
779 chain fatty acid levels in farmed Atlantic salmon, 2006-2015. *Scientific Reports*, 6, 21892.

780 Sprague, M., Walton, J., Campbell, P.J., Strachan, F., Dick, J.R., Bell, J.G. (2015).
781 Replacement of fish oil with a DHA-rich algal meal derived from *Schizochytrium* sp. on the
782 fatty acid and persistent organic pollutant levels in the diets and flesh of Atlantic salmon
783 (*Salmo salar* L.) post-smolts. *Food Chemistry*, 185, 413-421.

784 Tocher, D.R. (2015). Omega-3 long-chain polyunsaturated fatty acids and aquaculture in
785 perspective. *Aquaculture*, 449

786 Tocher, D.R., Betancor, M.B., Sprague, M., Olsen, R.E., Napier, J.A. (2019). Omega-3 long-
787 chain polyunsaturated fatty acids, EPA and DHA: Bridging the gap between supply and
788 demand. *Nutrients*, 11, 89.

789 Tocher, D.R., Harvie, D.G. (1988). Fatty acid compositions of the major phosphoglycerides
790 from fish neural tissues: (n-3) and (n-6) polyunsaturated fatty acids in rainbow trout (*Salmo*

791 *gairdneri* L.) and cod (*Gadus morhus*) brains and retinas. *Fish Physiology and*
792 *Biochemistry*, 5, 229-239.

793 Torstensen, B.E., Frøyland, L., Lie, Ø. (2004). Replacing dietary fish oil with increasing levels
794 of rapeseed oil and olive oil – effects on Atlantic salmon (*Salmo salar* L.) tissue and
795 lipoprotein lipid composition and lipogenic enzyme activities. *Aquaculture Nutrition*, 10,
796 175-192.

797 Turchini, G.M., Ng, W.-K., Tocher, D.R. (2010). *Fish oil replacement and alternative lipid*
798 *sources in aquaculture feeds* (1st ed) (p. 551). Boca Raton, Florida, USA: CRC Press.

799 Turchini, G.M., Torstensen, B.E., Ng, W.-K. (2009). Fish oil replacement in finfish nutrition.
800 *Reviews in Aquaculture*, 1, 10-57.

801 WHO. (2003). World Health Organization (WHO) Technical Report No. 916. *Diet, Nutrition*
802 *and the Prevention of Chronic Diseases*. WHO, Geneva, Switzerland.

803 Ytrestøyl, T., Aas, T.S., Åsgård, T. (2015). Utilisation of feed resources in production of
804 Atlantic salmon (*Salmo salar*). *Aquaculture*, 448, 365-374.

Figure Legends

Figure 1. Fatty acid compositions (% of total lipid, mean \pm SD) of the various salmon products surveyed with respect to a) 18:1n-9, b) 18:2n-6, c) 18:3n-3 and d) EPA+DHA. Samples are of Scottish farmed origin unless otherwise stated. Bars bearing different lettering are significantly different (ANOVA, $P < 0.05$) ($n = 3$). *Note, Retailer 2 sold two farmed Norwegian salmon products with different nutritional labelling, differing in the number of fillets per pack, 2 or 3 (Standard and Standard L, respectively).

Figure 2. Content of total EPA+DHA (g.100g flesh⁻¹ ww) in farmed salmon products available in the UK and separated based on farmed origin, Scotland and Norway respectively. Mean (---), median (—), interquartile range (box) and 10th and 90th percentiles (whiskers). Boxplots bearing different lettering are significantly different (ANOVA, $P < 0.05$). Fish from Retailer 4 was included in overall value but excluded elsewhere due to unknown origin ($n = 60, 42$ and 15 , Overall, Scotland and Norway respectively).

Figure 3. Absolute amounts of EPA+DHA in 130 g servings (mean \pm SD) of the various farmed salmon products surveyed in the present study. Samples are of Scottish farmed origin unless otherwise stated. Bars bearing different lettering are significantly different (ANOVA, $P < 0.05$) ($n = 3$). *Note, Retailer 2 sold two farmed Norwegian salmon products with different nutritional labelling, differing in the number of fillets per pack, 2 or 3 (Standard and Standard L, respectively).

Figure 4. Amount of EPA+DHA (g) per GBP (£) spent of farmed Atlantic salmon products surveyed in the present study. Samples are of Scottish farmed origin unless otherwise stated. Bars bearing different lettering are significantly different (ANOVA, $P < 0.05$) ($n = 3$). *Note, Retailer 2 sold two farmed Norwegian salmon products with different nutritional labelling, differing in the number of fillets per pack, 2 or 3 (Standard and Standard L, respectively).

Figure 5. Principal component analysis of the farmed salmon products surveyed in the present study based on farmed origin and measured nutrient variables (lipid and protein content and fatty acid and mineral composition) in farmed Atlantic salmon products of Scottish (●), Norwegian (▲) and unknown (×) farmed origin.

Figure 6. Comparison of the EPA+DHA contents (g.100g flesh⁻¹ ww) of salmon products from distinct farmed origins but containing the same nutritional labelling. *Note, Retailer 2 sold two farmed Norwegian salmon products with different nutritional labelling, differing in the number of fillets per pack, 2 or 3 (Standard and Standard L, respectively). Bars from products within the same retailer bearing different lettering are significantly different (ANOVA, $P < 0.05$) ($n = 3$).

Table 1. List of the salmon products sampled in the current study with respect to product range (value, standard, premium/organic), farmed origin (Scotland and Norway), price (£ per kg) and nutritional content as stated on product label. Retailers which stocked only one range was deemed as their standard product, irrespective of retailer status.

Retailer	Product Range ^a	Farmed Origin	Pack size (g) ^b	Price (£ per kg) ^c	Nutritional Content (per 100 g) ^d			
					Protein	Fat	Omega-3	EPA+DHA
1	Standard	Scotland	240	9.96	20.0	11.0	-	-
	Premium	Scotland	280	14.54	20.0	11.0	1.4	-
2	Standard	Scotland	240	11.04	23.0	17.0	-	2.7
	Standard	Norway	240	11.04	23.0	17.0	-	2.7
	Standard L ^d	Norway	340*	11.03	23.0	15.0	-	0.8
3	Standard	Scotland	220	18.18	19.0	14.0	3.1	-
4	Standard	Unknown ^e	520*	11.54	24.2	13.1	-	-
5	Standard	Scotland	240	9.96	22.0	19.0	3.6	-
6	Standard	Scotland	230	19.57	20.3	13.4	3.4	2.6
7	Standard	Norway	240	16.67	19.2	15.7	-	-
8	Value	Scotland	343	11.00	23.5	14.9	1.4	0.7
	Standard	Scotland	240	13.75	23.5	14.9	1.9	1.0
	Premium	Scotland	240	20.83	23.6	14.1	2.3	1.2
9	Standard	Scotland	270	11.11	23.6	14.2	2.4	-
	Standard	Norway	270	11.11	23.6	14.2	2.4	-
10	Value	Scotland	600*	15.53	19.1	15.6	1.9	-
	Standard	Scotland	280	24.96	19.1	15.6	1.9	-
	Organic	Scotland	265	26.38	18.8	13.6	1.5	-
Branded	Standard	Norway	240	16.67	23.0	14.7	-	0.8
	Organic	Scotland	252	22.00	23.5	13.9	-	1.8

^aProduct range defined by retail product label; retailers selling only one product range, which was not organic, considered as standard product.

^bPack size based on product label and contained two fillet portions, with exception to products marked * which contained a minimum of 3 fillet portions per pack.

^cPrice correct at time of purchase and excludes any promotional offers

^dAccording to product label. Values stated based on cooked (grilled, pan-fried, oven cooked) or uncooked conditions.

^eRetailer 2 contained 2 Norwegian standard products with different nutritional labelling and number of fillets per pack – defined as standard and standard L.

^fExact origin unknown, labelled as farmed in Scotland/Norway

848 **Table 2.** Mineral compositions of farmed Atlantic salmon products surveyed in the present study. Results are presented on a wet weight (ww)
849 basis.

	Retailer 1		Retailer 2			Retailer 3	Retailer 4	Retailer 5	Retailer 6	Retailer 7
	Standard	Premium	Standard	Standard	Standard L*	Standard	Standard	Standard	Standard	Standard
	Scotland	Scotland	Scotland	Norway	Norway	Scotland	Unknown [†]	Scotland	Scotland	Norway
Macro minerals g.kg ⁻¹ ww										
Sodium	0.31 ± 0.08 ^d	0.36 ± 0.08 ^{cd}	0.35 ± 0.07 ^{cd}	0.36 ± 0.02 ^{cd}	0.32 ± 0.03 ^d	0.33 ± 0.03 ^d	0.35 ± 0.04 ^{cd}	0.30 ± 0.04 ^d	0.27 ± 0.02 ^d	0.40 ± 0.12 ^{cd}
Potassium	3.34 ± 0.02	3.34 ± 0.14	3.54 ± 0.17	3.24 ± 0.38	3.58 ± 0.27	3.62 ± 0.27	3.18 ± 0.04	3.18 ± 0.20	3.35 ± 0.17	3.31 ± 0.17
Calcium	0.22 ± 0.05 ^{a-e}	0.24 ± 0.03 ^{a-d}	0.25 ± 0.07 ^{a-c}	0.29 ± 0.04 ^a	0.27 ± 0.01 ^{a-c}	0.29 ± 0.02 ^{ab}	0.19 ± 0.05 ^{b-f}	0.25 ± 0.03 ^{a-c}	0.18 ± 0.04 ^{c-f}	0.14 ± 0.03 ^{e-h}
Magnesium	0.24 ± 0.02	0.22 ± 0.00	0.24 ± 0.02	0.22 ± 0.01	0.23 ± 0.01	0.24 ± 0.01	0.22 ± 0.01	0.22 ± 0.01	0.22 ± 0.00	0.23 ± 0.02
Phosphorus	2.10 ± 0.05	2.05 ± 0.05	2.14 ± 0.11	1.97 ± 0.19	2.13 ± 0.10	2.18 ± 0.15	1.96 ± 0.05	1.97 ± 0.10	2.08 ± 0.07	2.03 ± 0.15
Trace elements mg.kg ⁻¹ ww										
Iron	4.13 ± 1.23 ^{a-c}	3.69 ± 0.77 ^{a-c}	2.88 ± 0.60 ^{a-c}	3.44 ± 0.83 ^{a-c}	4.65 ± 1.03 ^a	4.01 ± 0.75 ^{a-c}	2.21 ± 0.56 ^{a-c}	2.01 ± 0.35 ^{bc}	2.64 ± 0.43 ^{a-c}	2.71 ± 0.19 ^{a-c}
Manganese	0.13 ± 0.03 ^{a-d}	0.13 ± 0.01 ^{a-d}	0.14 ± 0.00 ^{ab}	0.14 ± 0.00 ^{a-c}	0.14 ± 0.00 ^{a-c}	0.15 ± 0.00 ^a	0.10 ± 0.02 ^{b-f}	0.11 ± 0.01 ^{a-e}	0.10 ± 0.02 ^{b-f}	0.09 ± 0.00 ^{d-f}
Copper	0.85 ± 0.24 ^a	0.69 ± 0.13 ^{ab}	0.55 ± 0.03 ^{ab}	0.64 ± 0.05 ^{ab}	0.76 ± 0.09 ^a	0.45 ± 0.14 ^{ab}	0.29 ± 0.05 ^b	0.30 ± 0.06 ^b	0.31 ± 0.07 ^b	0.45 ± 0.32 ^{ab}
Zinc	6.32 ± 0.76 ^{ab}	6.14 ± 0.55 ^{a-d}	6.41 ± 0.80 ^{ab}	6.23 ± 0.52 ^{a-c}	6.02 ± 0.29 ^{a-e}	6.54 ± 0.57 ^a	5.06 ± 0.69 ^{a-f}	5.23 ± 0.78 ^{a-f}	4.98 ± 0.70 ^{a-f}	4.48 ± 0.68 ^{d-f}
Vanadium	0.01 ± 0.00 ^{b-e}	0.01 ± 0.00 ^{de}	0.01 ± 0.00 ^{b-e}	0.02 ± 0.00 ^a	0.01 ± 0.01 ^{b-e}	0.01 ± 0.00 ^{c-e}	0.02 ± 0.01 ^{ab}	0.01 ± 0.00 ^{b-e}	0.00 ± 0.00 ^{de}	0.01 ± 0.00 ^{b-e}
Selenium	0.17 ± 0.01 ^{cd}	0.12 ± 0.01 ^{de}	0.13 ± 0.01 ^{de}	0.09 ± 0.01 ^e	0.12 ± 0.00 ^{de}	0.14 ± 0.01 ^{de}	0.08 ± 0.01 ^e	0.13 ± 0.03 ^{de}	0.13 ± 0.02 ^{de}	0.10 ± 0.04 ^{de}

850 **Table 2 cont.**

Value Range	Retailer 8			Retailer 9		Retailer 10			Branded	
	Value	Standard	Premium	Standard	Standard	Value	Standard	Organic	Standard	Organic
	Scotland	Scotland	Scotland	Scotland	Norway	Scotland	Scotland	Scotland	Norway	Scotland
Macro minerals g.kg ⁻¹ ww										
Sodium	0.60 ± 0.08 ^{bc}	0.70 ± 0.26 ^a	0.89 ± 0.08 ^{ab}	0.35 ± 0.08 ^{cd}	0.36 ± 0.03 ^{cd}	0.44 ± 0.03 ^{cd}	0.30 ± 0.06 ^d	0.30 ± 0.03 ^d	0.34 ± 0.04 ^d	0.32 ± 0.02 ^d
Potassium	3.39 ± 0.14	3.25 ± 0.24	3.18 ± 0.20	3.61 ± 0.15	3.46 ± 0.14	3.37 ± 0.07	3.37 ± 0.30	3.58 ± 0.43	3.66 ± 0.21	3.46 ± 0.08
Calcium	0.15 ± 0.04 ^{e-g}	0.14 ± 0.04 ^{e-h}	0.15 ± 0.06 ^{d-g}	0.09 ± 0.01 ^{f-h}	0.11 ± 0.02 ^{f-h}	0.10 ± 0.02 ^{f-h}	0.05 ± 0.02 ^h	0.04 ± 0.02 ^h	0.07 ± 0.02 ^{gh}	0.07 ± 0.00 ^{gh}
Magnesium	0.23 ± 0.01	0.23 ± 0.01	0.22 ± 0.01	0.23 ± 0.01	0.24 ± 0.01	0.22 ± 0.01	0.24 ± 0.03	0.23 ± 0.02	0.25 ± 0.04	0.24 ± 0.01
Phosphorus	2.12 ± 0.07	2.11 ± 0.10	2.00 ± 0.09	2.16 ± 0.09	2.15 ± 0.08	2.01 ± 0.05	2.14 ± 0.17	2.14 ± 0.18	2.25 ± 0.19	2.19 ± 0.06
Trace elements mg.kg ⁻¹ ww										
Iron	2.84 ± 1.11 ^{a-c}	1.70 ± 0.20 ^c	2.36 ± 0.59 ^{a-c}	1.92 ± 0.68 ^{bc}	2.34 ± 0.38 ^{a-c}	4.38 ± 1.86 ^{ab}	2.84 ± 1.12 ^{a-c}	2.63 ± 0.37 ^{a-c}	3.43 ± 0.52 ^{a-c}	3.21 ± 0.45 ^{a-c}
Manganese	0.09 ± 0.01 ^{c-f}	0.10 ± 0.02 ^{b-f}	0.07 ± 0.02 ^{ef}	0.08 ± 0.01 ^{ef}	0.08 ± 0.02 ^{ef}	0.07 ± 0.03 ^{ef}	0.07 ± 0.01 ^{ef}	0.07 ± 0.00 ^{ef}	0.06 ± 0.00 ^f	0.06 ± 0.01 ^f
Copper	0.46 ± 0.05 ^{ab}	0.68 ± 0.21 ^{ab}	0.59 ± 0.21 ^{ab}	0.31 ± 0.06 ^b	0.36 ± 0.03 ^b	0.48 ± 0.03 ^{ab}	0.49 ± 0.11 ^{ab}	0.48 ± 0.07 ^{ab}	0.57 ± 0.05 ^{ab}	0.34 ± 0.01 ^b
Zinc	4.80 ± 0.55 ^{b-f}	4.55 ± 0.56 ^{c-f}	4.35 ± 0.62 ^{ef}	4.27 ± 0.05 ^f	4.42 ± 0.09 ^{d-f}	4.55 ± 0.39 ^{c-f}	4.83 ± 0.45 ^{a-f}	4.48 ± 0.46 ^{d-f}	4.33 ± 0.34 ^{ef}	4.69 ± 0.54 ^{b-f}
Vanadium	0.01 ± 0.00 ^{c-e}	0.01 ± 0.00 ^{de}	0.01 ± 0.00 ^{b-d}	0.01 ± 0.00 ^{b-d}	0.01 ± 0.00 ^{a-c}	0.01 ± 0.00 ^{a-d}	0.01 ± 0.00 ^{b-e}	0.00 ± 0.00 ^e	0.00 ± 0.00 ^{de}	0.00 ± 0.00 ^{de}
Selenium	0.16 ± 0.02 ^{c-e}	0.16 ± 0.05 ^{c-e}	0.15 ± 0.01 ^{c-e}	0.15 ± 0.03 ^{c-e}	0.10 ± 0.00 ^{de}	0.13 ± 0.02 ^{de}	0.26 ± 0.06 ^{ab}	0.23 ± 0.02 ^{bc}	0.13 ± 0.04 ^{de}	0.32 ± 0.00 ^a

851 Means (± standard deviation) bearing different superscript lettering within the same row are significantly different (ANOVA, *P* < 0.05) (*n* = 3 samples per product)

852 *Retailer 2 contained 2 Norwegian products with different nutritional labelling and number of fillets per pack – defined as Standard and Standard L (2 and 3 fillet packs, respectively)

853 [†]Exact origin unknown, labelled as farmed in Scotland or No

Figure 1

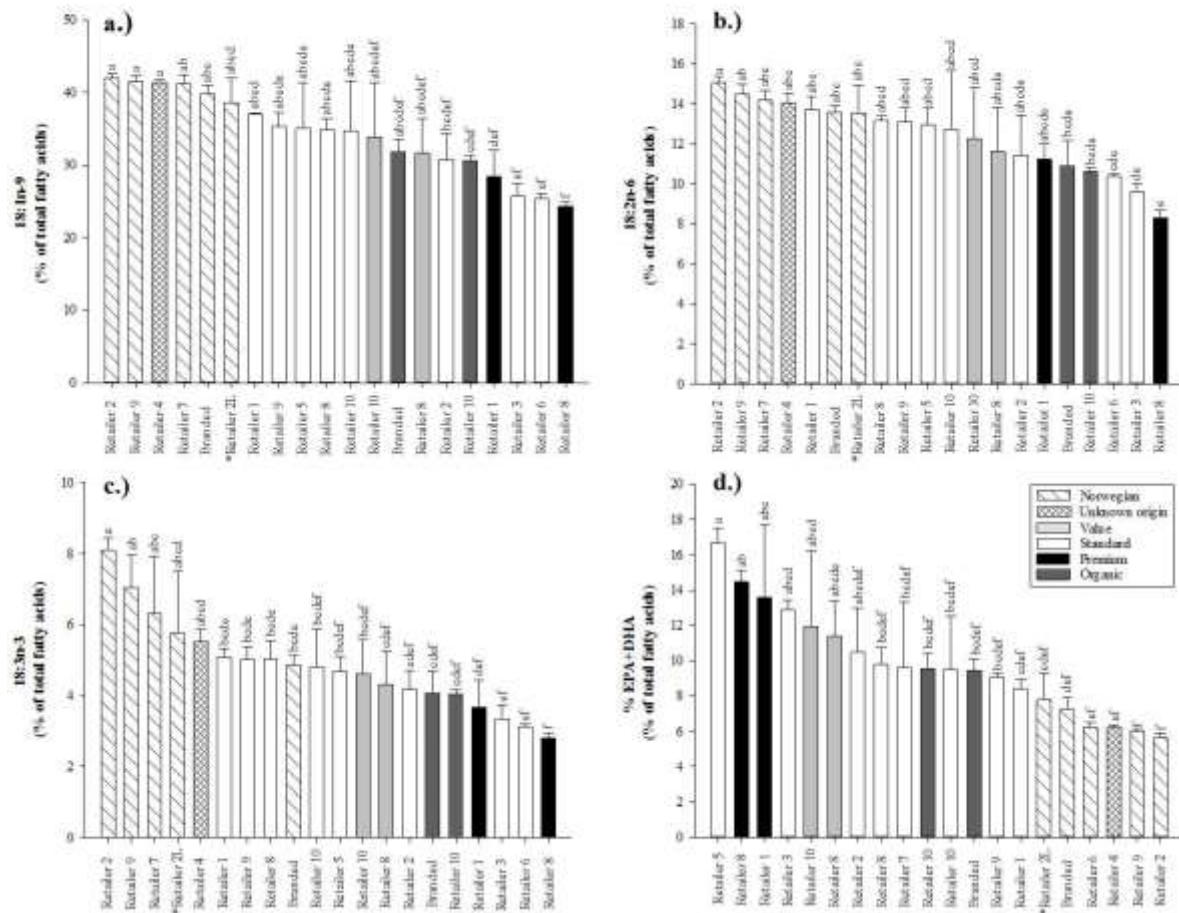
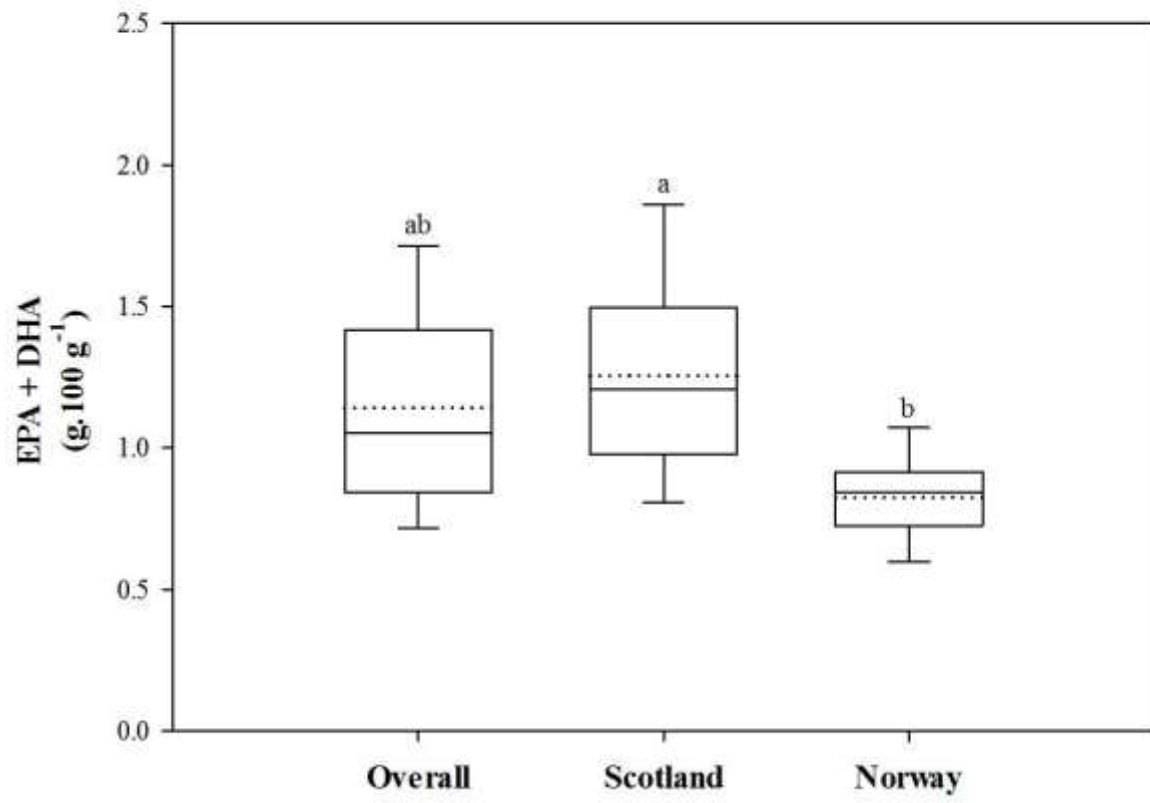
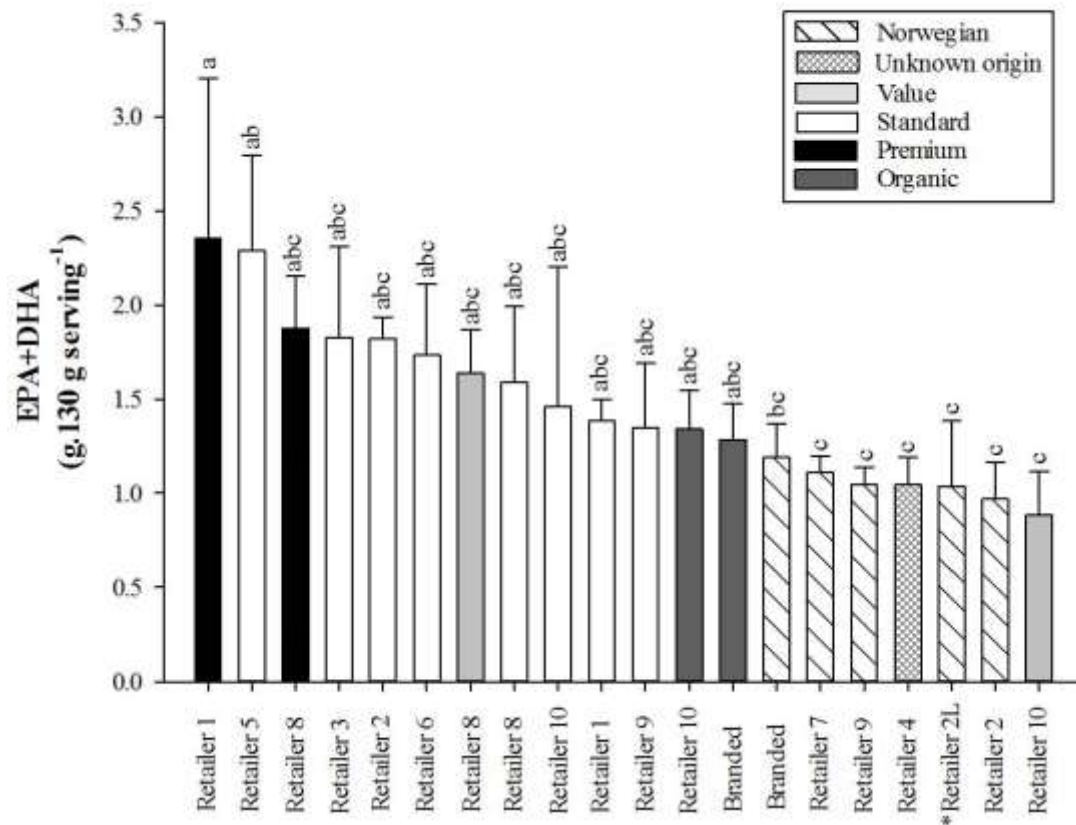


Figure 2



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Figure 3



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Figure 4

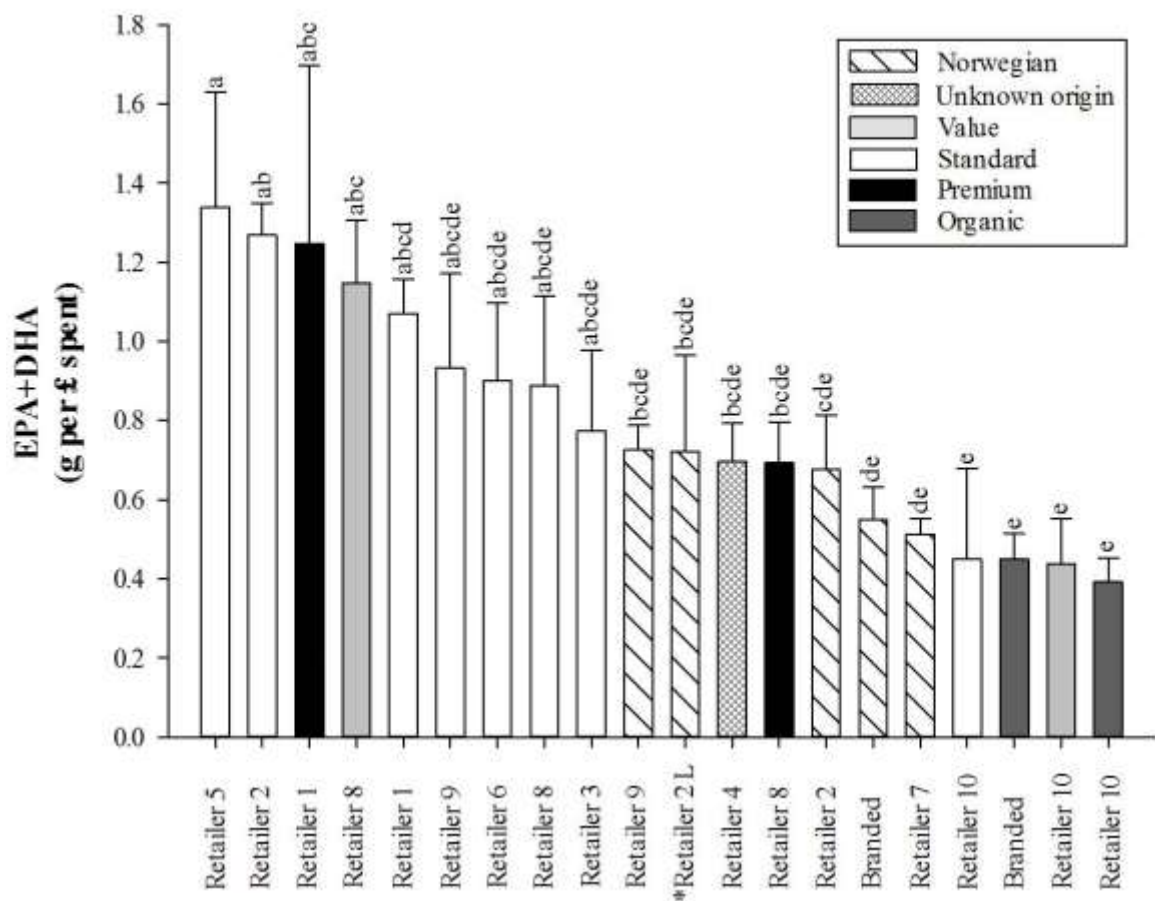


Figure 5

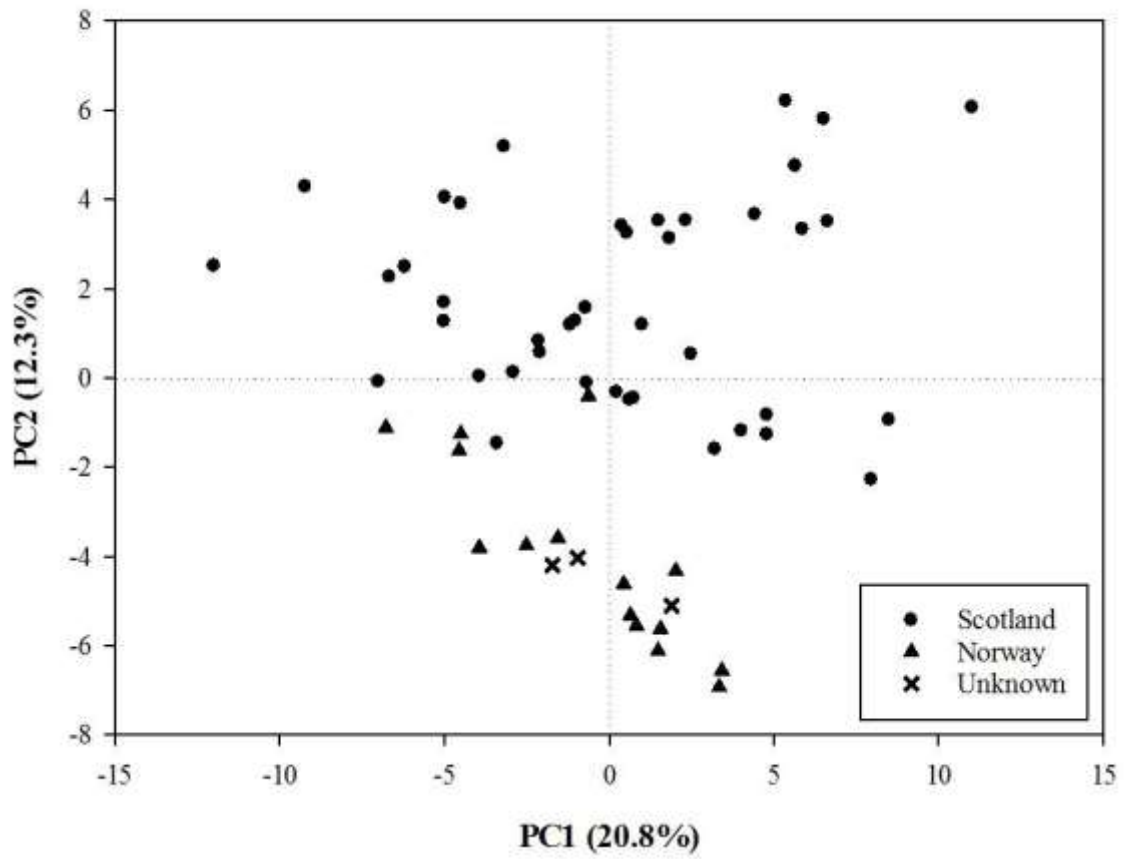


Figure 6

